ЕКОЛОГИЯ И КАЧЕСТВО НА ЖИВОТИНСКАТА ПРОДУКЦИЯ

ALTERNATIVES TO SURGICAL CASTRATION OF PIGS⁺

MARJETA ČANDEK-POTOKAR^{1,2}, NINA BATOREK LUKAČ¹

¹Agricultural Institute of Slovenia, Hacquetova ulica 17, SI-1000 Ljubljana, Slovenia ²University of Maribor, Faculty of Agriculture and Life Sciences, Pivola 10, SI-2311 Hoče, Slovenia

Castration or gonadectomy is a surgical procedure performed on male pigs for centuries in which testis and epididymis are physically removed from scrotal sack, without anaesthesia or analgesia, in the first days of life (Council **Directive**, 2008), mainly to get calmer and fatter pigs that do not exhibit boar taint and to prevent sexual behaviour in group housing (EFSA, 2004). For conventional production in the European Union (EU), castration is performed on 80 to 100% of male pigs (with the exception of Ireland 0%, United Kingdom 2.1%, Portugal 11.2%, Spain 33.3%, Cyprus 39% and Greece 75.6%), 97% of which are castrated without the use of anaesthesia (EFSA, 2004). Although generally used, this form of castration has been recently criticized as being painful for the piglets (McGlone et al., 1993 and Prunier et al., 2006) and may be considered even unnecessary in modern pig production with the emergence of new, alternative techniques, especially if pigs are raised for meat consumption where raising entire males (EM) would be more efficient (Bonneau, 1998 and Trefan et al., 2013). Moreover, the payment system of pig carcasses in most European countries rewards leanness, which gives EM additional advantage over surgical castrates (SC), provided that the carcasses are not being de-valorised due to the presence of boar taint. Thus, taking into account the welfare as well as the economic aspects it has been decided at the EU level that pig producers will voluntary stop castrating until 2018 (http://ec.europa.eu/food/ animal/welfare/farm/initiatives en.htm; http://boars2018. com) and at the same time, intensive research of alternative methods to surgical castration, diagnostic methods for boar taint analysis and harmonization of analytical methods is being supported.

Boar taint

Boar taint refers to the offensive odour or taste, evident during cooking or eating of meat or meat products from some, mostly EM pigs (**Bonneau et al.**, 1992). It makes pork undesirable for sensitive consumers (**Weiler et al.**, 2000). The substances that are considered the primary cause of boar taint are androstenone, a male pheromone, (**Patterson**, 1968) and skatole, a by-product of tryptophan breakdown in large intestine (**Vold**, 1970 and **Walstra and Maarse**, 1970). Androstenone (5α -androst-16-ene-3-one) is an anabolic steroid, without androgenic effect (**Claus et al.**, 1970), but with pronounced urine-like odour and flavour. It is produced by the Leydig cells in the testis of sexually mature male pigs (Gower, 1972). Due to its lipophilic character, it accumulates in adipose tissue in much higher amounts than other sex hormones (Pearce et al., 1988). Part of androstenone is stored in salivary glands, bound to specific binding protein pheromaxein (Booth and White, 1988), and after being released in the saliva, it serves as a pheromone to promote sexual behaviour in female pigs (Pearce et al., 1988). Unlike androstenone, skatole (3-metyl-indole) does not seem to have a biological function in pig. It is a byproduct of amino acid L-tryptophan breakdown by bacterial microflora in large intestine of pig. Thus the amount of skatole produced is primarily regulated by the availability of L-tryptophan which mainly originates from gut-mucosa cell debris (Claus et al., 1994 and Claus and Raab, 1999) and the activity of intestinal bacteria, especially Escherichia coli, Lactobacillus sp., Clostridium sp. (Jensen et al., 1995a) and Olsenella sp. (Jensen, 2012). Skatole that is produced in the colon is then partly excreted with faeces and partly absorbed through intestinal wall and released into the bloodstream. The metabolism and degradation of skatole in liver is inhibited by steroid hormones (Doran et al., 2002), and due to its lipophilic nature, it also accumulates in adipose tissue. The odour of skatole is described as faecal like or sometimes naphthalene like and can be detected by the vast majority of people (Weiler et al., 2000). On the contrary, androstenone has a large variation in consumer's sensitivity, as some people can detect it in very low concentrations, whereas approximately 25% are anosmic to it (Wysocki and Beauchamp, 1984 and Claus et al., 1994).

Alternatives to surgical castration of pigs

Possible alternatives to surgical castration with explanations of their way of action and the positive and negative aspects of their use and application are summarized in Table 1. Among them the alternatives that are the most promising are genetic selection for reduced boar taint, sex preselection (by sperm sorting) and rearing only females, immunocastration, surgical castration with pain relief (the use of analgesia/anaesthesia) and rearing of EM. Although one of the easiest ways to neutralize male fertility would be using substances that lead to local destruction of testicular tissue (e.g. formaldehyde, zinc or silver salt, acetic acid), because they are easy to administer, not expensive and

⁺ Статията е докладвана на научна конференция на ЗИ – Шумен "Иновации в аграрната наука за ефективно земеделие", организирана със съдействието на Министерството на образованието и науката през 2015 г.

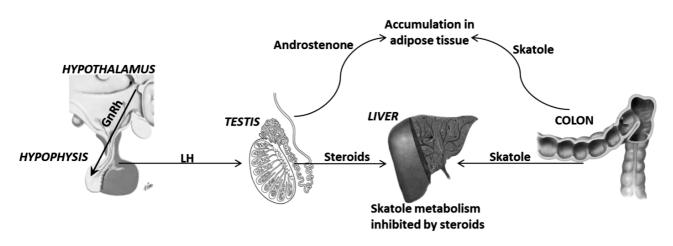


Fig. 1. Relationships between the hypothalamic-pituitary-gonadal axis, androstenone production in testes and skatole formation from tryptophan in intestine and their metabolism in liver. In boar, the production of testicular steroids, including androstenone, primarily a male pheromone, inhibits hepatic metabolism of skatole. Androstenone and skatole are therefore, due to their lipophilic character, deposited in adipose tissue.

cause no haemorrhages, the use of such substances causes inflammatory reaction, leading to swelling, necrosis and subsequently pain (Fahim, 1994 and EFSA, 2004) and is therefore unacceptable from a welfare point of view. Also the use of endogenous hormones (gonadotropin releasing hormone - GnRH agonists or antagonists) in order to neutralize GnRH provides no alternative to surgical castration in the EU, because their use is strictly limited to medical purposes, mostly for the treatment of reproductive disorders (Councile Directive, 1996 and Stephany, 2001). Thus, from a practical point of view only immunocastration, rearing of EM and castration with pain relief are acceptable alternatives in the EU, which can be applied immediately, whereas sex preselection by semen sorting and genetic selection for low androstenone level are long term, but sustainable and welfare friendly solutions (EFSA, 2004).

Castration with pain relief

Castration under anaesthesia or/and with the use of analgesia is basically the same surgical procedure as is the one being performed nowadays, except that with the application of pharmacological agents to the piglets prior or/and during the operation the pain is omitted or reduced. Hence, the use of analgesia/anaesthesia improves animal welfare of piglets significantly, but it generates extra costs on pig farms which depend on the substances being used and the form of their application. The costs increase significantly if the anaesthesia cannot be administered by farmers and when the veterinarian has to be called (de Roest et al., 2009). There are numerous possibilities of reducing or avoiding pain caused by castration that can be in general divided in local or general anaesthesia and analgesia alone, however they all require additional handling of the piglets inducing additional stress and greatly extending the time required for performing the castration (EFSA, 2004). The method of choice should result in a significant reduction or elimination of pain, not only sedation, discomfort and stress

for the piglets; moreover the substance has to be approved for the use in pigs reared for meat consumption (under Council Regulation No 2377/90), the dose of the substance has to be consistently adhered, because the toxic or even lethal dose can easily be exceeded in young piglets, and it would be preferable that it can be administered by the farmer alone, which significantly narrows down the available options. The remaining options, giving satisfactory results in term of pain relief, registered for use in animals raised for meat consumption, which might be used without additional equipment on the farm are:

 use of pre-operative analgesia alone – intra-muscular administration of non-steroidal anti-inflammatory drug (e.g. Ketoprofen, Meloxica);

- use of combination of pre-operative analgesia and local anaesthesia - intra-muscular administration of of non-steroidal anti-inflammatory drug and intratesticular or intrafunicular administration of 1% lidocaine with adrenalin.

Other options all have some drawbacks; the application of anaesthetic mixture via nasal spray is insufficient to induce adequate depth of anaesthesia (Axiak et al., 2007), general anaesthesia by injection was insufficient in significant number of piglets and has to be administered by veterinarian (Leeb et al., 2008), general anaesthesia by inhalation (isoflurane or sevoflurane - none is currently registered for the use in pigs) requires special equipment, veterinarians' assistance plus application of analgesic (e.g. non-steroidal anti-inflammatory drug), because it does not provide sufficient pain relief when used alone (Kupper et al., 2008; Schulz, 2007; Hodgson, 2007 and EFSA, 2004). There is a thin line between lethal and anaesthetic dose and also inhalation anaesthesia and CO₂ are aversive prior to the loss of consciousness (EFSA, 2004). Norway and Switzerland have already prohibited surgical castration without pain relief and have introduced alternative methods - the castration is performed by veterinarians and anaesthesia is mandatory (EFSA, 2004).

Table 1. Sumr	mary of alternativ	e methods to surg	Fable 1. Summary of alternative methods to surgical castration with explanations of their way of action and its pro et contra	ns of their way of action a	nd its pro et contra	
Alternative			Way of action	Pro	Contra	Reference
Raising entire male pigs	 Detection of boar Trained taint panellis slaughte 	r Trained panellists on slaughter line	Cooking test	Detecting tainted carcasses regardless to their source	Useful only on small number of animals, subjective method	Bundesanzeiger ,2007
			Soldering iron applied to the exposed backfat of the carcass	On line method for boar taint detection	Possibility of false positive or false negative results, subjective	Jarmoluk et al.,1970
		Skatole analysis on slaughter line	Spectrophotometric method	Successful and relatively fast (180 samples/hour)	Androstenone is not detected	Mortensen and Sorensen, 1984
		Electronic nose	Chemical electronic sensors which measure androstenone	<	Only experimental, 16% false positive	Annor-Frempong et al., 1998
			and skatole	occasion, all tainted carcasses are detected	results	Ampuero and Bee, 2006
		Mass spectrometry	Pyrolysis-mass spectrometry	High classification rates	Experimental	Ampuero and Bee , 2006 Ampuero et al., 2008
	Slaughter at		Slaughter of pigs below 90 kg	Reduces the incidence of Requires detection	Requires detection	Bonneau ,1987
	lower weight			tainted carcasses, higher % lean meat. better feed	of boar taint on slaughterline, lighter	Zamaratskaia et al., 2005 Aldal et al., 2005
					carcasses are less	Chen et al., 2007
					profitable, meat is unsuitable for dry-meat products	
	Controlling		Farrow-to-finish system,	Reduced skatole level.	No effect on	Hansen et al., 1994
	boar taint		feeding diets rich in indigestible	no adverse effect on	androstenone level	Jensen et al.,1995
	through nutrition	_	carbohydrates, clean	growth		Rideaut et al., 2004
	and rearing		environment, ideal temperature and max ventilation rate			Zamaratskaia et al., 2005 Andersson et al 2005
Blocking	Targeting GnRH	GnRH agonists	Negative feedback of GnRH		Short-term effect.	Reid et al., 1996
reproductive))	agonist on GnRH synthesis		hormonal treatment	Sinclair et al., 2001
axis		GnRH	Permanent occupation of GnRH		is expensive and not	Bruessow et al., 2011
		antagonists	receptors by antagonist		permitted in EU	Ziecik et al., 1989
		Passive immunization	Application of anti GnRH serum	Immunization effect in 24 h	Short-term effect, large amounts of antisera and	Van der Lende et al., 1993
					frequent administration is required	
		GnRH targeted toxins	Destruction of GnRH receptors bearing cells by GnRH coupled to evtotoxins	Single treatment reduces testosterone to 0 for approximately 20 weeks	Experimental, not used on pigs	Sabeur et al., 2003 Harrison et al., 2004

СЕЛСКОСТОПАНСКА АКАДЕМИЯ • ЖИВОТНОВЪДНИ НАУКИ, LII, 5/2015

43

Alternative			Way of action	Pro	Contra	Reference
	Targeting androstenone	Immunization against 5α- androst-16-en- 3-one	Ig that are produced neutralize androstenone in circulation	Reduction of androstenone conc. in adipose tissue, advantages of male type performance, carcass composition is preserved	Experimental	Williamson et al., 1985
Surgical castration	With local anaesthesia		Intrafunicular and/or intratesticular application of local anaesthetic (commonly lidocaine with adrenalin)	Reduces the acute pain and stress induced by castration	No economic benefits, additional cost, requires veterinary assistance, pain is not eliminated	EFSA, 2004 Prunier et al., 2006 Fredricksen and Nafstad, 2006
	With general anaesthesia	By injection	Intramuscular application of anaesthetic mixture prior castration	Reduction of acute pain	Long recovery period, losses are 3-5 %, insufficient depth of anaesthesia, additional cost and veterinary assistance	Lahrmann et al., 2006 Leeb et al., 2008 Zankl, 2007
		By inhalation	Anaesthesia with isouflurane by Reduction of pain mask	Reduction of pain	Expensive, time consuming, requires veterinarian assistance, high mortality of piglets	Shulz, 2007 Kupper et al., 2008
			CO2 inhalation	Fast introduction and recovery	Exposure to CO2 is aversive until loss of consciousness, possibility of overdose	Svendersen et al., 2005 Kluivers-Poodt et al., 2007
		By nasal spray	Administration of anaesthetic mixture (Ketamin, Azeperone and Climazolame) via nasal spray	Induction in 10 min and fast recovery, reduction of pain	Insufficient depth of anaesthesia, possibility of overdose	Axiak et al., 2007
	With analgesia	Meloxicam/ Metamizol / Ketoprofen	Application of analgesic prior castration	Effective pain relief	Additional cost and veterinary assistance is required	Heinritzi et al., 2006 Zöls et al., 2006
Chemical castration		Formaldehyde, zinc or silver salt, acetic acid	Local destruction of testicular tissue	Easy to administer, not expensive and no haemorrhages	Painful inflammatory reaction	Fahim, 1994 Giri et al., 2002

Alternative		Way of action	Pro	Contra	Reference
Sperm sexing	Sperm sexing Sperm Sexing Technology	Sex preselection (producing female litters) by flow- cytometry cell sorting	Females do not exhibit boar taint	To slow for commercial Johnson et al ., 1989 use, requires intrauterine Johnson , 2000 insemination, 85-95% purity, less efficient production and fatter carcasses than boars	Johnson et al., 1989 Johnson, 2000
	Semen sexing kit (high volume sperm sexing technology)	Binding x-bearing cells by Ig, filtration deagglutination of X- bearing cells	Faster than BSST, produces high- volume of x-bearing sperm (good potential for commercial use)	Experimental, 70% purity	ter Beek, 2007
Genetic selection	For low androstenone	Identifying genes responsible It would be the most for low androstenone welfare friendly method, production or high androstenone which would not require and skatole metabolism and castration or boar taint selection of such animals detection on line		It is a long term solution, Willeke et al., 1987 still in development Bonneau et al., 198 Willeke and Pirchn 1989 Sellier et al., 2000 Moe et al., 2007	Willeke et al., 1987 Bonneau et al., 1987 Willeke and Pirchner, 1989 Sellier et al., 2000 Moe et al., 2007

Raising entire males

Raising EM (boars) is a common practice in some EU countries (United Kingdom, Ireland, Spain, Portugal) and is gradually being introduced by some pig chains in the Netherlands, Germany, Belgium and France. Compared with SC rearing of EM it is more cost effective (no cost and wound infections due to castration, better FCR and better carcass conformation - higher meat percentage; Bonneau, 1998) .However, this type of farming is more demanding for the farmer and more stressful for the animals (more aggression and sexual behaviour as they mature; **Prunier et al.**, 2006); moreover there is a possibility for occurrence of boar taint in meat and meat products from EM (rejected by majority of consumers; Malmfors and Lundström, 1983 and Weiler et al., 2000), lower meat quality (decreased tenderness and lower water holding capacity, reduced intramuscular fatness; Babol and Squires, 1995 and Trefan et al., 2013) and subsequently decreased suitability for processing into high quality meat products (Diestre et al., 1990 and Banon et al., 2003). Because the occurrence of boar taint at commercial slaughter weights is very variable, ranging from 10 to 75% (EFSA, 2004), depending on breed, slaughter weight, rearing and management conditions, the production of EM requires screening and sorting of carcasses on the slaughter line. Currently there are only subjective methods that can be applied (cooking test and use of soldering iron applied on exposed backfat of the carcass; Bundesanzeiger, 2007 and Jarmoluk et al., 1970) that require trained panellist, as the objective methods that would give immediate or quick result are either still experimental (electronic nose and mass spectrometry; Annor-Frempong et al., 1998 and Ampuero and Bee, 2006) or detect only one component of boar taint (spectrophotometric method for skatole analysis and ELISA kit for androstenone analysis; Mortensen and Sornsen, 1984 and Walstra et al., 1999). In recent years, several strategies to prevent or reduce occurrence of boar taint meat have been developed. They include the use of special feeding strategies (different non-starch polysaccharides and additives; Albrecht, 2011), slaughter at a lower weight (prior sexual maturity; Bonneau, 1987) and improving rearing conditions (farrow-to-finish system, clean environment, maximal ventilation rate and temperature in termoneutral zone; Salmon and Edwards; 2006, Hansen et al., 1994 and Andersson et al., 2005). Thus, if detection of the tainted meat on the slaughter line and improved processing of tainted meat is provided, the rearing of EM can easily be applied.

Immunocastration

Immunocastration uses natural immune system of the pig for the formation of specific antibodies that bind and neutralize GnRH. It consists of two vaccinations, as a result hypothalamic-pituitary-gonadal axis is blocked and testes growth and sexual steroids synthesis are effectively inhibited (see Fig. 2). Physiologically the immunocastration becomes effective in a week following second vaccination (V2; **Claus et al.,** 2007), therefore in case of late revaccination during the month preceding slaughter, growth characteristics

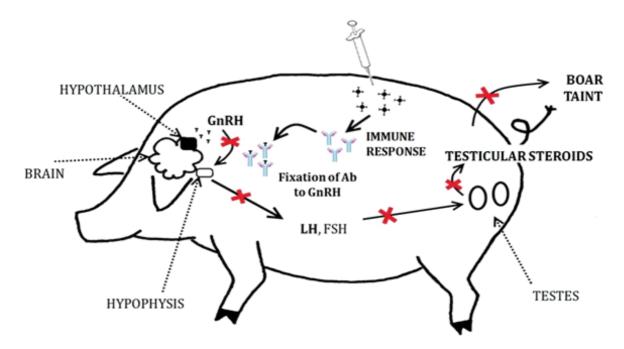


Fig. 2 Physiological response to immunocastration in male pig: after revaccination a large quantity of antibodies against GnRH are produced that neutralise GnRH. As GnRH is no longer available, there is no stimulus for follicle-stimulating hormone and luteinizing hormone release from hypophysis and spermatogenesis and production of androgen hormones in testis is on hold, meaning that there is neither semen production nor male behaviour.

of immunocastrated pigs (IC) are similar to those of EM (Škrlep et al., 2010; Millet et al., 2011 and Dunshea et al., 2013) and better production performance (i.e. growth rate, feed intake, feed efficiency and carcass leanness) of IC compared with SC (for review see Millet et al., 2011 and Batorek et al., 2012) is observed. The only currently available commercial vaccine - Improvac[®] (Zoetis Florham Park, NJ, USA) was introduced on the market in Australia and New Zealand in the nineties, and is now registered for use in more than 50 countries around the world, including the EU (since 2009). Although the vaccine producer recommends that a 4-week interval between V2 and slaughter is respected, recent studies (Lealiifano et al., 2011 and Kubale et al., 2013) indicated that the concentration of androstenone and skatole in fat tissue drop below the limit of sensory detection already two weeks after V2 in pigs of commercial slaughter weight. Simultaneously with the decrease of steroid hormones and boar taint compounds concentrations, the effect of immunocastration is visible on the reproductive organs through the reduced size of the testes and accessory sex glands (Bonneau, 2010), with the strongest impact on the size of vesicular gland, followed by the size of the bulbourethral gland. Regression of the reproductive tract is consistent with the loss of functional activity as shown by histological changes of testicular tissue (i.e. atrophy of Leydig cells and germinal epithelium, the cessation of spermatogenesis, the reduction of the size of glandular acini and the disappearance of secretory function of accessory sex glands; Fang et al., 2010; Einarsson et al., 2011 and Kubale et al., 2013). Although studies suggest that the effect of immunocastration persisted up

to 22 weeks (Zamaratskaia et al., 2008) it should not be permanent (Claus et al., 2008). However, an irreversible loss of reproductive capacity can occur following the earlier vaccination protocol (Einarsson et al., 2009). Regarding the effectiveness of immunization, it should be noted that some animals do not react to it (so-called "non-responders") due to poor immunological response or technically improper vaccination (Zeng et al., 2002; Jaros et al., 2005 and Škrlep et al., 2012). The number of non-responders is relatively low (1-3%) and is expected to be similar to the number of cryptorchids obtained after surgical castration. Thus, there is a need of a fast, simple but effective method for boar taint detection on the slaughter line also when rearing IC. Due to the large variability in size of testes among animals (also due to differences in age and weight), the evaluation of the effectiveness of immunocastration on slaughter line based on the size of testes is not reliable and the size of the accessory sex glands should be take into account as well (Bonneau, 2010 and Čandek-Potokar et al., 2014), which is rather unpractical.

Sex preselection

The only currently available accurate and potentially cost effective approaches for achieving sex preselection is separating X- from Y-chromosome-bearing sperm with high volume sperm sexing technology (sperm sexing kit; **Ter Beek**, 2007) or on the basis of the difference in electric charge by using modified flow-cytometer/cell sorter system (**Johnson et al.**, 1989). Cell sorting system can produce about 18 million X-sperm per h of 85-90% of purity (**Johnson**, 2000) whereas semen sorting kit results in 70% purity, which gives the possibility of breeding exclusively female offspring. However, both methods are technologically under developed and currently unsuitable for commercial use in the pig (Hofmo, 2006), as the boar semen is rather sensitive to manipulation compared to other animal species and consequently relatively large insemination dose is required (3 billion spermatozoa in 80-100 mL of fresh diluted semen; Dominiek et al., 2011). However, using post-cervical or deep intrauterine insemination allows 3 to 20 fold reduction in the number of spermatozoa in the insemination dose compared to the standard transcervical artificial insemination (Vazquez et al., 2008), but special skills are needed for catheter handling and there is a possibility of damaging cervical or uterine tissue (Dominiek et al., 2011). Controlled-release capsules containing 2.5 billion boar spermatozoa per dose were developed to extend the preservation time of spermatozoa and maximize the efficiency of a single artificial insemination (Vigo et al., 2009) in swine. Thus, sperm sexing and insemination with X-sperm have a great potential in pig production provided that the reproductive performance is optimized by the use of accurate and precise insemination technique and that semen sorting results in a larger quantity of high quality semen, that would meet the market requirements.

Genetic selection for low androstenone level

Genetic selection for low androstenone level is another long-term and welfare friendly solution that is still in development, although selection of breeds that have a natural low incidence of boar taint could provide short-term solution (EFSA, 2004). Sexually mature boars that have low fat androstenone but normal plasma testosterone and luteinizing hormone values have been identified almost 30 years ago (Bonneau, 1987). However, single nucleotide polymorphisms associated with boar taint compounds that can be applied in practical breeding to reduce boar taint in intact boars without aversive effect on sexual maturity have been identified only recently (Moe et al., 2009) and require further research before being implemented in practice.

CONCLUSION

If surgical castration is associated with pain and risk to the health of animals, non-castration is associated with social stress and fighting occurring when boars reach puberty, resulting in skin lesions and ultimate carcass damage. It is more profitable to raise EM pigs due to their enhanced feed conversion ratio and greater carcass leanness; however the quality of meat from boars may be deteriorated by its odour or taste, lower intramuscular fat content and reduced tenderness. Thus, raising EM and applying castration when they approach puberty would mean combining benefits of the two alternatives; benefiting from boar performance during most of the growth-finish period, while avoiding the risk of boar taint and aggression among pigs. This becomes possible with the use of late immunocastration. However, according to current indications it seems that for fresh meat consumption rearing of entire males will be practiced, whereas immunocastration remains a good alternative in the case of fattening to a greater weight and also, although commercially less important for culled boars.

REFERENCES

1. Albrecht, A., 2012. Growth performance and carcass characteristics of Improvac TM -treated male pigs compared with barrows. Doctoral thesis. University of Veterinary Medicine Hannover, , 33 pp.

2. Aldal, I., Ø. Andresen, A.K. Egeli, J.E. Haugen, A. Grødum, O. Fjetland and J.L.H. Eikaas, 2005. Levels of androstenone and skatole and the occurrence of boar taint in fat from young boars. Livest. Prod. Sci., 95, 1: 121-129.

3. Ampuero, S., M. Amrhein, S. Dubois, and G. Bee, 2008. Identification of boar off-flavour with an electronic nose. In: Proceedings of the EAAP meeting, Girona, Spain. pp.1-2.

4. Ampuero, S. and G. Bee, 2006. The potential to detect boar tainted carcasses by using an electronic nose based on mass spectrometry. Acta Agr. Scan., 48, Suppl I: P1.

5. Andersson, K. H., K. Andersson, G. Zamaratskaia, L. Rydhmer, G. Chen and K. Lundström, 2005. Effect of single-sex or mixed rearing and live weight on performance, technological meat quality and sexual maturity in entire male and female pigs fed raw potato starch. Acta Agr. Scan., Section A-Animal Science, 55, 2-3: 80-90.

6. Annor-Frempong, I. E., G. R. Nute, J. D. Wood, F. W. Whittington and A. West, 1998. The measurement of the responses to different odour intensities of boar taint using a sensory panel and an electronic nose. Meat Sci., 50: 139–151.

7. Axiak, S. M., N. Jäggin, S. Wenger, M. G. Doherr and U. Schatzmann, 2007. Anaesthesia for castration of piglets: Comparison between intranasal and intramuscular application of ketamine, climazolam and azaperone. Schweiz. Arch. Tierheilk., 149, 9: 395-402.

8. Babol, J., E.J. Squires and K. Lundström, 1998. Relationship between oxidation and conjugation metabolism of skatole in pig liver and concentrations of skatole in fat. J. Anim. Sci., 76: 829-838.

9. Banon, S., E. Costa, M. D. Gil and, M. D. Garrido, 2003. A comparative study of boar taint in cooked and dry-cured meat. Meat Sci., 63, 3: 381-388.

10. Batorek, N., M. Čandek-Potokar, M. Bonneau and J. Van Milgen, 2012. Meta-analysis of the effect of immunocastration on production performance, reproductive organs and boar taint compounds in pigs. Animal, 6, 8: 1330-1338.

11. Bonneau, M., 1987. Effects of age and live weight on fat 5α -androstenone levels in young boars fed two planes of nutrition. Reprod. Nutr. Dev., 27, 2A: 413-22.

12. Bonneau, M., M. Le Denmat, J. C. Vaudelet, J. R. Veloso Nunes, A. B. Mortensen and H. P. Mortensen, 1992. Contributions of fat androstenone and skatole to boar taint: I. Sensory attributes of fat and pork meat. Livest. Prod. Sci., 32, 1: 63-80.

13. Bonneau, M., N. Meusy-Dessolle, P. C. Lüglise and R. Claus, 1982. Relationships between fat and plasma androstenone and plasma testosterone in fatty and lean young boars following castration. Acta Endocrinol., 101: 129-133.

14. Bonneau, M., 1998. Use of entire males for pig meat in the European Union. Meat Sci., 49: 257-272.

15. Bonneau, M., 2010. Accessory sex glands as a tool to measure the efficacy of immunocastration in male pigs. Animal, 4: 930-932.

16. Booth, W.D. and White C.A., 1988. The isolation, purification and some properties of pheromaxein, the pheromonal steroid-binding protein, in porcine submaxillary glands and saliva. J. Endocrinol., 118, 1: 47-NP.

17. Bruessow, K.P., F. Schneider, K. Wollenhaupt and A. Tuchscherer, 2011. Endocrine effects of GnRH agonist application to early pregnant gilts. J. Reprod. Dev., 57, 2: 242-248.

18. Bundesanzeiger, 2007. Allgemeine Verwaltungsvorschriftüber die Durchfürung der amtlichen Überwachung der Einhaltung von Hygienvorschriften für Lebensmittel tierischen Ursprungs und zum Verfahren zur Prüfung von Leitlinien für eine gute Verfahrenspraxis (AVV Lebenmittelhygiene — AVV LmH). Jhg 59, no 180a: 51–52. (Ger.)

19. Čandek-Potokar, M., M. Prevolnik and M. Skrlep, 2014. Testes weight is not reliable tool for discriminating immunocastrates from entire males. V: POPOVIĆ, Zoran (ur.). Proceedings of the International Symposium on Animal Science 2014, 23-25th September 2014, Belgrade, Serbia. Belgrade Faculty of Agriculture, 2014, str. 43-49.

20. Chen, G., E. Bourneuf, S. Marklund, G. Zamaratskaia, A. Madej and K. Lundström, 2007. Gene expression of 3β -hydroxysteroid dehydrogenase and 17β -hydroxysteroid dehydrogenase in relation to androstenone, testosterone, and estrone sulphate in gonadally intact male and castrated pigs. J. Anim. Sci., 85, 10: 2457-2463.

21. Claus, R., B. Hoffmann and H. Karg, 1970. Determination of 5α -androst-16-en-3-one and testosterone in peripheral plasma and testicular tissue of pigs. In: Third International Congress on Hormonal Steroids Hamburg Germany 1970, Abstract 49.

22. Claus, R., M. Lacorn, K. Danowski, M.C. Pearce, A. Bauer, 2007. Short-term endocrine and metabolic reactions before and after second immunization against GnRH in boars. Vaccine, 25: 4689-4696.

23. Claus, R. and S. Raab, 1999. Influences on skatole formation from tryptophan in the pig colon. In Tryptophan, Serotonin, and Melatonin, Springer US. pp. 679-684.

24. Claus, R., U. Weiler and A. Herzog, 1994. Physiological aspects of androstenone and skatole formation in the boar – a review with experimental data. Meat Sci., 38: 289-305.

25. Council Directive, 2008. Council Directive 2008/120/EC of 18 December 2008 laying down minimum standards for the protection of pigs.

26. de Roest, K., C. Montanari, T. Fowler and W. Baltussen, 2009. Resource efficiency and economic implications of alternatives to surgical castration without anaesthesia. Animal, 3: 1522-1531.

27. Diestre, A., M. A. Oliver, M. Gispert, I. Arpa and J. Arnau, 1990. Consumer responses to fresh meat and meat products from barrows and boars with different levels of boar taint. Anim Prod., 50, 3: 519-530.

28. Dominiek, M., A. Lopez Rodriguez, T. Rijsselaere, P. Vyt and A. Van Soom, 2011. Artificial Insemination in Pigs, Artificial Insemination in Farm Animals, Dr. Milad Manafi (Ed.), ISBN: 978-953-307-312-5, InTech, DOI: 10.5772/20630. Retrived from: http://www.intechopen.com/ books/artificial-insemination-in-farm-animals/artificialinsemination-in-pigs (accessed 16.3.2015)

29. Doran, E., F. W. Whittington, Wood J. D., J. D. McGivan, 2002. Cytochrome P450IIE1 (CYP2E1) is induced by skatole and this induction is blocked by androstenone in isolated pig hepatocytes. Chem Biol Interact, 140: 81-92.

30. Dunshea, F. R., J. R. D. Allison, M. Bertram, D. D. Boler, L. Brossard, R. Campbell, J. P. Crane, D. P. Hennessy, L. Huber, C. de Lange, N. Ferguson, P. Matzat, F. McKeith, P. J. U. Moraes, B. P. Mullan, J. Noblet, N. Quiniou and M. Tokach, 2013. The effect of immunization against GnRF on nutrient requirements of male pigs: A review. Animal, 7: 1769-1778.

31. EFSA, 2004. Welfare Aspects of the Castration of Piglets. Scientific Report of the Scientific Panel for Animal Health and Welfare, European Food Safety Authority 2004. EFSA J., 91: 1-19.

32. Einarsson, S., K. Andersson, M. Wallgren, K. Lundström, H. Rodriguez-Martinez, 2009. Short- and long-term effects of immunization against gonadotropinreleasing hormone, using Improvac[™], on sexual maturity, reproductive organs and sperm morphology in male pigs. Theriogenology, 71: 302-310.

33. Einarsson, S., C. Brunius, M. Wallgren, K. Lundström, K. Andersson, G. Zamaratskaia, H. Rodriguez-Martinez, 2011. Effects of early vaccination with Improvac[®] on the development and function of reproductive organs of male pigs. Anim. Reprod. Sci., 127: 50-55.

34. Fahim, M. S., 1994. U.S. Patent No. 5, 372, 822. Washington, DC: U.S. Patent and Trademark Office.

35. Fang, F., H. Li, Y. Liu, Y. Zhang, Y. Tao, Y. Li, H. Cao, S. Wang, L. Wang, X. Zhang, 2010. Active immunization with recombinant GnRH fusion protein in boars reduces both testicular development and mRNA expression levels of GnRH receptor in pituitary. Anim. Reprod. Sci., 119: 275-281.

36. Fredriksen, B. and O. Nafstad, 2006. Surveyed attitudes, perceptions and practices in Norway regarding the use of local anaesthesia in piglet castration. Res. Vet. Sci., 81: 293–295.

37. Giri, S. C., B. P. S. Yadav and S. K. Panda, 2002. Chemical castration in pigs. Indian J. Anim. Sci., 72: 451-453.

38. Gower, D. B., 1972. 16-Unsaturated C 19 steroids. A review of their chemistry, biochemistry and possible physiological role. J. Steroid Biochem., 3: 45-103.

39. Hansen, L. L., A. E. Larsen, B. B. Jensen, J. Hansen-Mller and P. Barton-Gade, 1994. Influence of stocking rate and faeces deposition in the pen at different temperatures on skatole concentration (boar taint) in subcutaneous fat. Anim. Prod., 59: 99-110.

40. Harrison, G. S., M. E. Wierman, T. M. Nett and

L. M. Glode, 2004. Gonadotropin-releasing hormone and its receptor in normal and malignant cells. Endocr-relat. Cancer, 11: 725-748.

41. Heinritzi, K., S. Zöls and M. Ritzmann, 2006. Possibilities of pain-reduction in castration of piglets. In: Proceedings of the 19th International Pig Veterinary Society Congress Copenhagen 2006, p 289.

42. Hodgson, D. S., 2007. Comparison of isoflurane and sevoflurane for short-term anesthesia in piglets. Vet. anaesth. analg., 34: 117-124.

43. Hofmo, P. O., 2006. Sperm sorting and low dose insemination in the pig – an update. Acta Vet Scand., 48: 11.

44. Jarmoluk, L., H. A. Martin, H. T. Freeden, 1970. Detection of taint (sex odour) in pork. Can. J. Anim. Sci., 50: 750.

45. Jaros, P., E. Bbrgi, K.D.C. Stärk, R. Claus, D. Hennessy, R. Thun, 2005. Effect of immunization against GnRH on androstenone concentration, growth performance and carcass quality in intact male pigs. Livestock Livest. Prod. Sci., 92: 31-38.

46. Jensen, M. T., R. P. Cox and B. B. Jensen, 1995. 3-Methylindole (skatole) and indole production by mixed populations of pig fecal bacteria. Appl. Environ. Microb., 61: 3180-3184.

47. Jensen, R. L., 2012. Feed interventions and skatole deposition. Masters thesis, University of Copenhagen.

48. Johnson, L. A., J. P. Flook and H. W. Hawk, 1989. Sex preselection in rabbits: live births from X and Y sperm separated by DNA and cell sorting. Biol. Reprod., 41: 199–203.

49. Johnson, L.A., 2000: Sexing mammalian sperm for production of offspring: the state-of-the-art. Anim. Reprod. Sci., 60–61: 93–107.

50. Kluivers-Poodt, M., H. Hopster and H. A. M. Spoolder, 2007. Castration under anaesthesia and/or analgesia in commercial pig production. Animal Science Group Wageningen 2007, Report 85.

51. Kubale, V., N. Batorek, M. Škrlep, A. Prunier, M. Bonneau, G. Fazarinc, M. Čandek-Potokar, 2013. Steroid hormones, boar taint compounds, and reproductive organs in pigs according to the delay between immunocastration and slaughter. Theriogenology, 79: 69-80.

52. Kupper, T., C. Pauly, C. Burren, A. Hofer and P. Spring, 2008. Alternative Methoden zur konventionellen Ferkelkastration ohne Schmerzausschaltung. Projekt ProSchwein Abschlussbericht, Schweizerische Hochschule für Landwirtschaft SHL Zollikofen 2008. (Ger.)

53. Lahrmann, K. H., M. Kmiec and R. Stecher, 2006. Die Saugferkelkastration mit der Ketamin/Azaperon-Allgemein anaesthesie: tierschutzkonform, praktikabel, aber wirtschaftlich? Prakt. Tierarzt., 87: 802–809. (Ger.)

54. Lealiifano, A. K., J. R. Pluske, R. R. Nicholls, F. R. Dunshea, R. G. Campbell, D. P. Hennessy, D. W. Miller, C. F. Hansen, B. P. Mullan, 2011. Reducing the length of time between harvest and the secondary gonadotropin-releasing factor immunization improves growth performance and clears boar taint compounds in male finishing pigs. J. Anim. Sci., 89: 2782-2792.

55. Leeb, C., C. Gößler, B. Czech and J. Baumgartner,

2008. Experiences with intravenous general anaesthesia for surgical castration of pigs. In Book of Abstracts of the 59th Annual Meeting of the EAAP, Vilnius. p. 105.

56. Malmfors, B., K. Lundström, 1983. Consumer reactions to boar meat-a review. Livest. Prod. Sci., 10, 2: 187-196.

57. McGlone, J. J., R. I. Nicholson, J. M. Hellman, D. N. Herzog, 1993. The development of pain in young pigs associated with castration and attempts to prevent castration-induced behavioral changes. J. Anim. Sci., 71: 1441-1446.

58. Millet, S., K. Gielkens, D. De Brabander, G. P. J. Janssens, 2011. Considerations on the performance of immunocastrated male pigs. Animal, 5: 1119-1123.

59. Moe, M., S. Lien, T. Aasmundstad, T. H. Meuwissen, M. H. Hansen, C. Bendixen and E. Grindflek, 2009. Association between SNPs within candidate genes and compounds related to boar taint and reproduction. BMC genetics, 10, 1: 32.

60. Moe, M., T. Meuwissen, S. Lien, C. Bendixen, X. Wang, L.N. Conley and E. Grindflek, 2007. Gene expression profiles in testis of pigs with extreme high and low levels of androstenone. BMC genomics, 8, 1: 405.

61. Mortensen, A. B. and S. E. Sorensen, 1984. Relationship between boar taint and skatole determined with a new analysis method. Proc. 30th European Mtg. of Meat Res. Workers, Bristol, U. K. Paper 8-11, p. 395.

62. Patterson, R. L. S., 1968. 5α-androst-16-ene-3-one: Compound responsible for taint in boar fat. J. Sci. Food Agr., 19: 31-38.

63. Pearce, G. P., P. E. Hughes, W. D. Booth, 1988. The involvement of boar submaxillary salivary gland secretions in boar-induced precocious puberty attainment in the gilt. Anim. Reprod. Sci., 16, 2: 125-134.

64. Prunier, A., M. Bonneau, E. H. von Borell, S.Cinotti , M. Gunn, B. Fredriksen, M. Giershing, D. B. Morton, F. A. M. Tuyttens and A. Velarde, 2006. A review of welfare consequences of surgical castration in piglets and the evaluation of non-surgical methods. Anim. Welfare, 15: 277-289.

65. Reid, J., J. J. Dufour and M. A. Sirard, 1996. Effect of a single injection of a long-acting gonadotropin-releasing hormone agonist on prepubertal male and female pigs on reproductive organs, growth performance and sensory qualities of pork roasts. Reprod. Nutr. Dev., 36: 321-332.

66. Rideout, T.C., M. Z. Fan, J. P. Cant, C. Wagner-Riddle and P. Stonehouse, 2004. Excretion of major odorcausing and acidifying compounds in response to dietary supplementation of chicory inulin in growing pigs. J. Anim. Sci., 82, 6: 1678-1684.

67. Sabeur, K., B. A. Ball, T. M. Nett, H. H. Ball and I. K. Liu, 2003. Effect of GnRH conjugated to pokeweed antiviral protein on reproductive function in adult male dogs. Reproduction, 125, 6: 801-806.

68. Salmon, E.L.R. and S.A. Edwards, 2006. Effects of gender contact on the behaviour and performance of entire boars and gilts from 60–130 kg. In: Proceedings of the British Society of Animal Science, York, UK, p. 72.

69. Schulz, C., 2007. Auswirkungen einer Isofluran-Inhalationsnarkose auf den Kastrationsstress und die postoperativen Schmerzen von Ferkeln. Dissertation, Ludwig-Maximilians-Universität München. pp.128. (Ger.)

70. Sellier, P., P. Le Roy, M. N. Fouilloux, J. Gruand and M. Bonneau, 2000. Responses to restricted index selection and genetic parameters for fat androstenone level and sexual maturity status of young boars. Livest. Prod. Sci., 63, 3: 265-274.

71. Sinclair, P. A., E. J. Squires, J. I. Raeside, J. H. Britt and V. G. Hedgpeth, 2001. The effect of early postnatal treatment with a gonadotropin-releasing hormone agonist on the developmental profiles of testicular steroid hormones in the intact male pig. J. Anim. Sci., 79, 4: 1003-1010.

72. Stephany, R. W., 2001. Hormones in meat: different approaches in the EU and in the USA. Apmis, **109**, 103: 357-364.

73. Svendsen, O., L. Strobeck and B. Forman, 2005. CO2/O2 as an anaesthetic agent during castration of piglets. In: EAAP working group on production and utilization of meat from entire male pigs. Uppsala, Sweden.

74. Škrlep, M., N. Batorek, M. Bonneau, G. Fazarinc, B. Šegula and M. Čandek-Potokar, 2012. Elevated fat skatole levels in immunocastrated, surgically castrated and entire male pigs with acute dysentery. Vet. J., 194, 3: 417-419.

75. Ter Beek, V., 2007. Sex sorting technology could save time. Pig progress, 23, 10: 23.

76. Trefan, L., A. Doeschl-Wilson, J. A. Rooke, C. Terlouw and L. Bünger, 2013. Meta-analysis of effects of gender in combination with carcass weight and breed on pork quality. J. Anim. Sci, 91, 3: 1480-1492.

77. Van der Lende, T., L. Kruijt and M. Tieman, 1993. Can passive immunization with anti-GnRH monoclonal antibodies, injected a few weeks before slaughter, prevent boar taint? In: Measurement and Prevention of Boar Taint, Ed. M. Bonneau, INRA Editions, Paris, p. 201-206.

78. Vazquez, J. M., J. Roca, M. A. Gil, C. Cuello, I. Parilla, J. L. Vazquez and E. A. Martínez, 2008. New developments in low-dose insemination technology. Theriogenology, 70: 1216 1224.

79. Vigo, D., M. Faustini, S. Villani, F. Orsini, M. Bucco, T. Chlapanidas, U. Conte, K. Ellis and M. L. Torre, 2009. Semen controlled-release capsules allow a single artificial insemination in sows. Theriogenology, 72: 439-444.

80. Vold, E., 1970. Fleishproduktionseigenschaften bei Ebern Kastraten IV: Organoleptische und gaschromatographische Untersuchungen wasserdampfflüchtiger Stoffe des Reckenspeckes von Ebern. Maldinger fra Norges Landbrukshøgskole, 49: 1-25. (Ger.)

81. Walstra, P. and G. Maarse, 1970. Onderzoek gestachlengen von mannelijke mestvarkens. IVO rapport,

Rapport 2 C-147. IVO rapport C-147. Researchroep voor Vlees en Vleeswaren TNO.

82. Walstra, P., C. Claudi-Magnussen, P. Chevillon, G. von Seth, A. Diestre, K.R. Matthiews, D.B. Homer, M. Bonneau, 1999. An international study on the importance of androstenone and skatole by country and season. Livest. Prod. Sci., 62: 15-28.

83. Weiler, U., M. Font i Furnols, K. Fischer, H. Kemmer, M.A. Oliver, M. Gispert, A. Dobrowolski, R. Claus, 2000. Influence of differences in sensitivity of Spanish and German consumers to perceive androstenone on the acceptance of boar meat differing in skatole and androstenone concentrations. Meat Sci., 54, 3: 297-304.

84. Willeke, H., R. Claus, E. Mbller, F. Pirchner and H. Karg, 1987. Selection for high and low level of 5α -androst-16-en-3-one in boars. J. Anim. Breed. Genet., 104, 1-5: 64-73.

85. Willeke, H. and F. Pirchner, 1989. Selection for high and low level of 5a-androst-16-en-3- one in boars. II. Correlations between growth traits and 5-androstenone. J. Anim Breed Genet., 106: 312–317.

86. Williamson, E. D., R. L. S. Patterson, E. R. Buxton, K.G. Mitchell, I. G. Partridge and N. Walker, 1985. Immunization against 5α -androstenone in boars. Livest. Prod. Sci., 12, 3: 251-264.

87. Wysocki, C. J. and G. K. Beauchamp, 1984. Ability to smell androstenone is genetically determined. In: Proceedings of the National Academy of Science of the USA vol. 81, pp. 4899–4902.

88. Zamaratskaia, G., J. Babol, H. K. Andersson, K. Andersson and K. Lundström, 2005. Effect of live weight and dietary supplement of raw potato starch on the levels of skatole, androstenone, testosterone and oestrone sulphate in entire male pigs. Livest. Prod. Sci., 93, 3: 235-243.

89. Zankl, A., 2007. Untersuchungen zur Wirksamkeit und Gewebeverträglichkeit von Lokalanästhetika bei der Kastration männlicher Saugferkel. Vet. med. Diss. Мъпсhen. pp.125. (Ger.)

90. Zeng, X. Y., J. A. Turkstra, R. H. Meloen, X. Y. Liu, F. Q. Chen, W. M. M. Schaaper, H. B. Oonk, D. Z. Guo and D. F. M. van de Wiel, 2002. Active immunozation against gonadotropin – releasing hormone on Chinese male pigs: effects of dose on antybody titer, hormone levels and sexual development. Anim. Reprod. Sci., 70: 223-233.

91. Ziecik, A. J., K. L. Esbenshade and J. H. Britt, 1989. Effects of a gonadotrophin-releasing hormone antagonist on gonadotrophin secretion and gonadal development in neonatal pigs. J. Reprod. Fert., 87: 281-289.

92. Zöls, S., M. Ritzmann and K. Heinritzi, 2006. Effect of local anaesthesia in castration of piglets. Tierärztliche Praxis Großtiere, 34: 103-106.

ALTERNATIVES TO SURGICAL CASTRATION OF PIGS+

M. Čandek-Potokar^{1,2}, N. Batorek Lukač¹

¹Agricultural Institute of Slovenia, Hacquetova ulica 17, SI-1000 Ljubljana, Slovenia ² University of Maribor, Faculty of Agriculture and Life Sciences, Pivola 10, SI-2311 Hoče, Slovenia

SUMMARY

Castration or gonadectomy is a surgical procedure performed on male pigs in which testis and epididymis are physically removed from scrotal sack, without anaesthesia or analgesia, in the first days of life, mainly to get calmer and fatter pigs that do not exhibit boar taint and to prevent sexual behaviour in group housing. Although generally used, this form of castration has been recently criticized as being painful for the piglets and may be considered even unnecessary in modern pig production with the emergence of new, alternative techniques, especially if pigs are raised for meat consumption where raising entire males would be more efficient. Thus, taking into account the welfare as well as the economic aspects it has been decided at the level of the European Union that pig producers will voluntary stop castrating until 2018. At the same time, intensive research of alternative methods to surgical castration, diagnostic methods for boar taint analysis and harmonization of analytical methods is being supported. This publication presents alternative methods to surgical castration of piglets without anesthesia or analgesia - their mode of action and the positive and the negative aspects of their application. According to current indications the most promising alternatives are immunocastration, surgical castration with analgesia or anesthesia, rearing of entire males, genetic selection on the reduced boar taint and seamen sorting technique and subsequently rearing of exclusively female offspring. The latter two methods are long-term, sustainable and animal welfare friendly, whereas the remaining three alternatives are suitable for immediate introduction into practice. It seems that for fresh meat consumption rearing of entire males will be practiced, whereas immunocastration remains a good alternative in the case of fattening to a greater weight.

Key words: pig, castration, immunocastration, welfare, entire males

Abbreviations: SC – surgical castrates, EM – entire males, IC – immunocastrated pigs, EU – European Union, GnRH – gonadotropin releasing hormone, V2 – second vaccination for immunocastration

⁺ This article was reported at a scientific conference of AI-Shumen "Innovations in agricultural science for effective agriculture", organized in collaboration with the Ministry of Education and Science in 2015.