

<https://doi.org/10.61308/VHQU9624>

# Fecal nitrogen excretion in dairy cows supplemented with feed additive containing active dry yeast

Vasil Vasilev\*, Evgeni Videv

Agricultural Academy, Agricultural Institute - Stara Zagora, Bulgaria

\*Corresponding author: vsvqualitet@abv.bg

Vasilev, V. & Videv, E. (2023). Fecal nitrogen excretion in dairy cows supplemented with feed additive containing active dry yeast. *Bulgarian Journal of Animal Husbandry*, 60(4), 29-35

## Abstract

The aim of the study was to determine the fecal excretion of N (TN and  $\text{NH}_4^+\text{-N}$ ) in dairy cows receiving the feed supplement Biolife® (Eurovix S.p.A., Italy) containing active dry yeast *Saccharomyces cerevisiae*. Two groups of 14 Holstein cows (B – experimental; C – control) at tie-stalled breeding participated in the experiment. Group B received the feed supplement through the total mixed ration (TMR). Fresh feces samples from both groups were collected on d 7, 14, 21, 28, 35 from the start of supplementation. Mean values of fecal pH and excreted DM did not differ between groups. The concentration of  $\text{NH}_4^+\text{-N}$  in the fecal samples of group B was reduced by 28.05%, average for the entire studied period ( $P<0.05$ ). In the period d 21-35, this reduction was 35% ( $P=0.095$ ). Excreted TN, averaged over the entire period, was reduced by 16.18% in group B, but the difference was not statistically significant. The reduction of TN content in feces from group B was more pronounced in the period d 21-35, when the reduction was 31.99% compared to group C ( $P=0.084$ ). A period of at least 14 days is required for adaptation to the supplement and for an occur effect on N excretion.

**Keywords:** fecal nitrogen excretion; dairy cows; active dry yeast; feed additive

## Introduction

Plant nitrogen (N), mostly in the form of protein, is an essential feed component for animal growth and development. Most of the consumed N is excreted, providing nutrients in the manure needed for crop growth. The problem with this cycling of N is that large losses usually occur, contributing to environmental pollution (Rotz, 2004). N losses and ammonia emissions from animal husbandry represent a significant water and air pollutant (Hristov, 2013). Besides increasing the agricultural land involved in manure use, the application of feeding and management techniques capable of increasing N retention efficiency and reducing N release per unit of animal product should be promoted (Xiccato et al., 2005). Improving N utilization by dairy cows and especially reducing N excretion in fecal

masses is desirable because of concerns about the contribution of agriculture to nitrogen pollution, especially ammonia (Kebreab et al., 2002). Reference values for N excretion from different livestock systems are required for the implementation of the Nitrate Directive (91/676/EC) (Xiccato et al., 2005).

Active dry yeast is a widely used feed additive in the dairy industry (Li et al., 2021), with varying success for favorably modifying the rumen environment and promoting microbial growth (Hristov et al., 2010). The yeast *Saccharomyces cerevisiae* stimulates microbial activity in the rumen, and thus more forage N is incorporated into the microbial fraction and the flow of microbial N reaching the small intestine is increased (Williams and Newbold, 1990; Williams et al., 1990; Karr et al., 1991; Erasmus et al., 1992; Newbold et al., 1995, 1998; Hristov et al., 2010; Tristant

and Moran, 2015). An increase in the total number of bacteria in the rumen as well as specific groups of microbes has been observed in a number of studies (Wiedmeier et al., 1987; Harrison et al., 1988; Dawson et al., 1990; Williams and Newbold, 1990; Erasmus et al., 1992; Newbold et al., 1998; Li et al., 2021). Greater bacterial biomass in the rumen will improve forage digestibility (Newbold et al., 1998), which may reduce urinary N losses (Hristov et al., 2010).

Supplementation with the yeast *Saccharomyces cerevisiae* reduced rumen ammonia concentration in some studies (Harrison et al., 1988; Erasmus et al., 1992; Piva et al., 1993; Enjalbert et al., 1999; Kamra et al., 2002; Alshaikh et al., 2002; Doležal et al., 2005; Moallem et al., 2009; Pinloche et al., 2013; Aoki et al., 2021). Lower ammonia concentrations are consistent with an observed increase of bacteria concentrations in rumen, which is associated with increased absorption of  $\text{NH}_3\text{-N}$  into microbial proteins and increased influx of bacterial N to the small intestine (Williams and Newbold, 1990; Erasmus et al., 1992; Hristov et al., 2010; McAllister et al., 2011). Microbial protein synthesis in rumen is an effective indicator of N utilization (Chen and Gomes, 1992; Dias et al., 2018). The concentration of  $\text{NH}_3\text{-N}$  in the rumen can be used to predict the efficiency of incorporation (absorption and retention) of dietary N into microbial protein (Bach et al., 2005), and therefore for studying feed efficiency.

A meta-analysis by Desnoyers et al. (2009) showed an increased digestibility of organic matter (OM) in whole tract and dry matter intake (DMI) in ruminants supplemented with *Saccharomyces cerevisiae*. In a number of experiments with cows supplemented with *S. cerevisiae*, crude protein (CP) digestibility was improved (Wiedmeier et al., 1987; Erasmus et al., 1992; Panda et al., 1995; Wohlt et al., 1991, 1998; Yoon and Stern, 1996; Miller-Webster et al., 2002; Lascano et al., 2012; Leicester et al., 2016; Perdomo et al., 2020; Li et al., 2021; Phe-satcha et al., 2022). Based on the mechanisms of action, described in the literature of *S. cerevisiae* supplements, we set ourselves the goal of determining nitrogen excretion in the feces of dairy

cows supplemented with the Biolife® feed additive containing active dry yeast *Saccharomyces cerevisiae*.

## Material and methods

For the purposes of the present study, fresh fecal samples obtained from an experimental dairy cows fed with the feed additive Biolife® (manufacturer: Eurovix S.p.A., Italy) were examined. The supplement contains dehydrated yeast of *Saccharomyces cerevisiae*, malted barley, soya lecithin, and flour of seaweed. According to the manufacturer's description (Eurovix: Biolife – Technical data sheet, *Mod. 416*), a continuative use of the product in the feed ration improves the functional balance of intestinal microflora, preventing formation of toxins that may alter the organic balance of the animal; more over the better using of the feed contribute to reduce the faecal emissions of undesired compounds (above all the ammonia) reducing the environmental impact and also the olfactive pollution in animal farms.

Two groups of Holstein cows (B – experimental; C – control) of 14 animals per group were kept tie-stalled until the end of the experiment. The Biolife® feed additive was added to the concentrate mixture at a level of 500 g/ton, and supplied to the experimental group of cows with the total mixed ration (TMR). Fresh feces samples from both groups were taken in the morning before feeding on days 7, 14, 21, 28 and 35 from the start of Biolife® supplementation. After rapid unification and homogenization of the individual samples, mid-samples were separated, sealed and de-aerated in freezer bags and delivered to a laboratory for pH, dry matter (DM), total N (TN) and ammonium N ( $\text{NH}_4^+\text{-N}$ ) analysis.

The pH values were determined after dilution, homogenization and stabilization of the samples with deionized water in a ratio of 1:2.5 (feces:water) and measured using a pH meter. Dry matter content (DM, %) was determined after drying in an dryer at 105 °C to constant weight. The ammonia complex ( $\text{NH}_4^+\text{-N}$ , mg/kg) was extracted with 2.0 mol/L KCl and subjected to colorimetric analysis (Nesslerization method).

Total nitrogen content (TN, %) in feces was determined by the Kjeldahl method (AOAC International, 2002). The obtained results were summarized using the statistical package MYSTAT 12 (SYSTAT Software, Inc., 2007).

## Results and discussion

Table 1 presents the results of the study, statistical analysis was performed for both periods whole experiment (d 7-35) and for the period from the 21st to the 35th day (d 21-35).

The mean pH values of the freshly excreted feces did not differ between the control (C) and

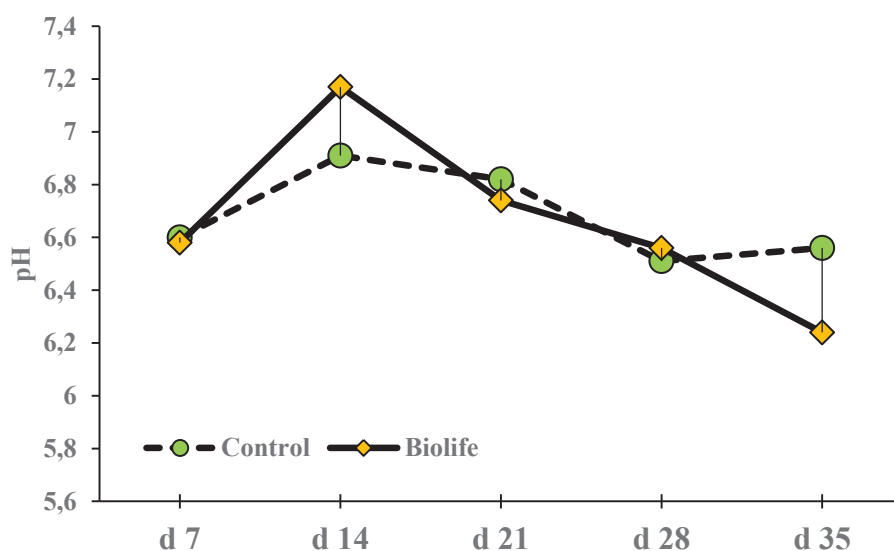
the experimental (B) group of cows, and they were 6.68 and 6.66, respectively, for the entire measurement period. For the period d 21-35, a slight decrease of 0.12 pH-units was reported in group B (Figure 1). The determined pH values of the feces were in the reference range for lactating cows (Palladino et al., 2022) with a daily variation of 5.80 to 6.76.

There were also no differences in DM content in fecal samples from groups C and B (on average 20.58% and 21.44%, respectively, for the whole period). However, in the period d 21-35 there was a more pronounced but insignificant increase in excreted dry matter in group B. Overall, the variation in fecal DM content was

**Table 1.** Effect of the feed additive Biolife® on fecal pH, DM, NH<sub>4</sub><sup>+</sup>-N, and TN in dairy cows (mean ± SD)

Variable	Period and treatment					
	d 7 to 35 (n=5)			d 21 to 35 (n=3)		
	C	B	P-value	C	B	P-value
pH	6.68±0.17	6.66±0.34	0.827	6.63±0.16	6.51±0.25	0.394
DM (%)	20.58±1.18	21.44±2.14	0.490	20.47±0.81	22.00±2.51	0.295
NH <sub>4</sub> <sup>+</sup> -N (mg/kg)	265.69±79.24	191.16±64.34	0.019*	239.67±89.54	155.77±58.87	0.095
TN (%)	0.686±0.040	0.575±0.165	0.232	0.695±0.050	0.472±0.103	0.084

C – Control; B - Biolife®; \* -  $P < 0.05$



**Fig. 1.** Dynamics of pH values in fresh excreted feces of dairy cows, supplemented with Biolife®

low, and within the range published by Weiss (2004) from 11.9% up to 20.9% in lactating Holstein cows.

The concentration of  $\text{NH}_4^+\text{-N}$  in fecal samples of group B (191.16 mg/kg) was reduced by 28.05% compared to that of group C (265.69 mg/kg), average over the entire study period ( $P < 0.05$ ). In the period d 21-35, this reduction was even higher (35.00%), however, it was not statistically significant ( $P = 0.095$ ). In general, for the entire experimental period,  $\text{NH}_4^+\text{-N}$  values had a moderately high degree of variation in both groups (CV=29.8% in group C and 33.7% in group B). Figure 2 reflects the dynamics in fecal concentrations of  $\text{NH}_4^+\text{-N}$ . In a study by Hristov et al. (2010), manure from dairy cows fed with a supplement of *Saccharomyces cerevisiae* fermentation product has a reduced ammonia emission potential.

Excreted total nitrogen (TN) were average 0.686% for group C and 0.575% for group B, over the entire experimental period, presenting a 16.18% reduction in TN in animals receiving Biolife®, but the difference was without statistical significance ( $P = 0.232$ ). TN excretion in group C maintained a constant level (Figure 3) with minimal variation (CV=5.8%), which may reflect constant daily dry matter intake (DMI)

and ration crude protein (CP) during the experiment, due to the established a positive correlation between daily fecal N excretion and individual variables such as DMI, N intake and ration CP content (Bougouin et al., 2022). The low variation in excreted DM (CV=5.8%) in group C may also be a result of constant dry matter intake.

The reduction of TN in fecal samples from group B was more pronounced in the period d 21-35 (Figure 3), when the reduction was 31.99% compared to group C, but the difference again did not reach statistical significance ( $P = 0.084$ ). It can be concluded that a minimum period of 14 days is necessary for animals to adapt to the supplement and to report an effect on N excretion. The values for excreted TN in our study overlap with those published by Brown (2013) for dairy cows, who from an analysis of 278 fecal samples (DM 18-30%) obtained a mean value of 0.69% for TN.

Lascano et al. (2012) observed a quadratic decrease in fecal N excretion with increasing daily dose of live yeast *S. cerevisiae* (0, 10, 30, and 50 g/d) in Holstein heifers, but urinary N excretion was similar in different doses. This resulted in no effect of live yeast on total excreted N. However, total-tract N apparent digestibil-

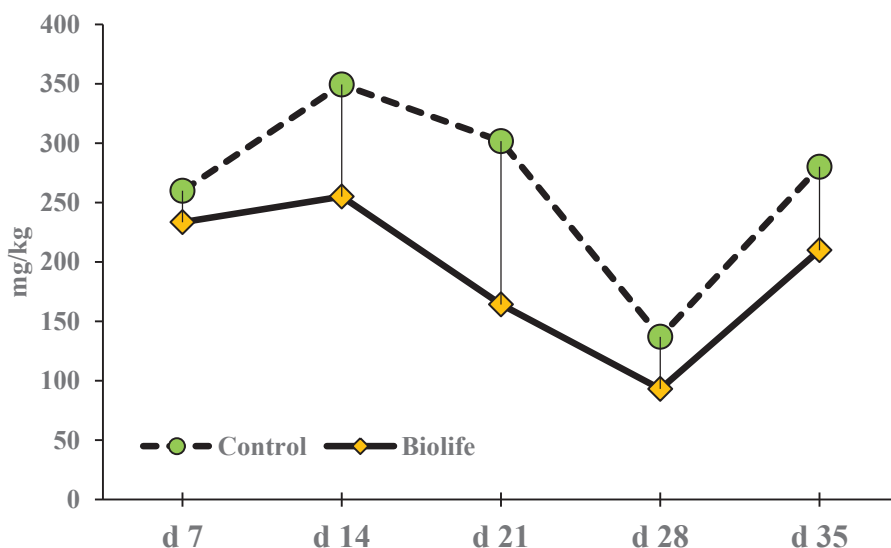


Fig. 2. Dynamics of  $\text{NH}_4^+\text{-N}$  in fresh excreted feces of dairy cows, supplemented with Biolife®

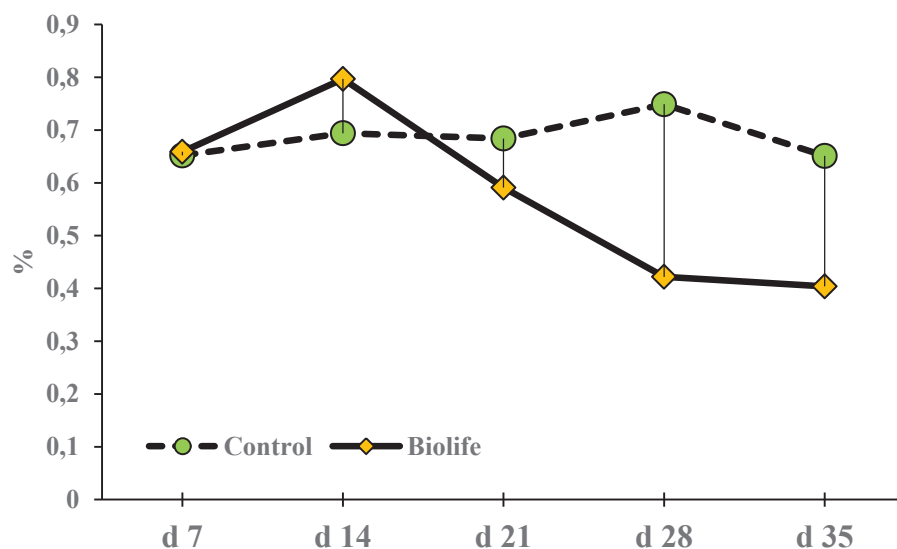


Fig. 3. Dynamics of TN in fresh excreted feces of dairy cows, supplemented with Biolife®

ity and retained N, expressed as a percentage of digested N, increased quadratically with increasing daily live yeast dose, reaching a maximum at 30 g/d. In a study by Chen et al. (2017), supplementation of Xiangzhong Black beef with *S. cerevisiae* significantly reduced fecal crude protein excretion, while Cunha et al. (2019) did not found effect of *S. cerevisiae* in Nellore heifers on N intake, fecal and urinary N excretion, N balance, urinary urea, N utilization efficiency (in relation to N intake and absorbed N), and microbial efficiency.

### Conclusions

Supplementation of dairy cows with the feed additive Biolife® reduced fecal  $\text{NH}_4^+\text{-N}$  concentration by 28.05% for the period d 7-35 ( $P < 0.05$ ) and by 35.00% for the period d 21-35 ( $P = 0.095$ ). The supplement reduced TN excreted in feces by 16.18% average for the entire study period, but without statistical significance, while in the period d 21-35 the reduction of TN was by 31.99% ( $P = 0.084$ ). Mean values of fecal pH and excreted DM did not differ between groups. It can be concluded that a period of at least 14 days is needed to adapt to the feed additive and show an effect on nitrogen excretion.

### References

- Alshaikh, M. A., Alsiadi, M. Y., Zahran, S. M., Mogawer, H. H. & Aalshowime, T. A. (2002). Effect of feeding yeast culture from different sources on the performance of lactating Holstein cows in Saudi Arabia. *Asian-Australian Journal of Animal Science*, 15(3), 352-356.
- AOAC International (2002). *Official methods of analysis*. 17<sup>th</sup> ed. 1<sup>st</sup> revision. AOAC Int., Gaithersburg, MD, USA.
- Aoki, N., Yanli, Z., Kanda, S., Kurokawa, Y., Sultana, H. & Itabashi, H. (2021). Effect of *Saccharomyces cerevisiae* fermentation product on ruminal fermentation, blood metabolites, and milk production in dairy cows. *JARQ*, 55(3), 265-271.
- Bach, A., Calsamiglia, S. & Stern, M. D. (2005). Nitrogen metabolism in the rumen. *Journal of Dairy Science*, 88(E. Suppl.), E9-E21.
- Bougouin, A., Hristov, A., Dijkstra, J., Aguerre, M. J., Ahvenjärvi, S., Arndt, C., Bannink, A., Bayat, A. R., Benchaar, C., Boland, T., Brown, W. E., Crompton, L. A., Dehareng, F., Dufrasne, I., Eugène, M., Froidmont, E., van Gastelen, S., Garnsworthy, P. C., Halmemies-Beauchet-Filleau, A., Herremans, S., Huhtanen, P., Johansen, M., Kidane, A., Kreuzer, M., Kuhla, B., Lessire, F., Lund, P., Minnée, E. M. K., Muñoz, C., Niu, M., Nozière, P., Pacheco, D., Prestløkken, E., Reynolds, C. K., Schwarm, A., Spek, J. W., Terranova, M., Vanhatalo, A., Wattiaux, M. A., Weisbjerg, M. R., Yáñez-Ruiz, D. R., Yu, Z. & Kebreab, E. (2022). Prediction of nitrogen excretion from data on dairy cows fed a wide range of diets compiled in an intercontinental data-

base: A meta-analysis. *Journal of Dairy Science*, 105(9), 7462-7481.

**Brown, C.** (2013). Available nutrients and value for manure from various livestock types. *Factsheet, Order No. 13-043*, AGDEX 538. Published by the Ontario Ministry of Agriculture and Food and the Ministry of Rural Affairs.

**Chen, L., Jie, H. D., Ren, A., Zhou, C. D., Tan, Z. L. & Li, B.** (2017). Effect of *Saccharomyces cerevisiae* on nutrient digestibility, rumen fermentation and plasma biochemical parameters of Xiangzhong Black beef. *Chinese Journal of Animal Nutrition*, 29, 3359-3365.

**Chen, X. B., & Gomes, M. J.** (1992). Estimation of microbial protein supply to sheep and cattle based on urinary excretion of purine derivatives. In: *An Overview of Technical Details; Occasional Publication*; International Feed Resources Unit, Rowett Research Institute: Aberdeen, Scotland, UK.

**Cunha, C. S., Marcondes, M. I., Silva, A. L., Gionbelli, T. R. S., Novaes, M. A. S., Knupp, L. S., Virginio Junior, G. F. & Veloso, C. M.** (2019). Do live or inactive yeasts improve cattle ruminal environment? *Revista Brasileira de Zootecnia*, 48, e20180259.

**Dawson, K. A., Newman, K. E. & Boling, J. A.** (1990). Effects of microbial supplements containing yeast and lactobacilli on roughage-fed ruminal microbial activities. *Journal of Animal Science*, 68, 3392-3398.

**Desnoyers, M., Giger-Reverdin, S., Bertin, G., Duvaux-Ponter, C. & Sauvant, D.** (2009). Meta-analysis of the influence of *Saccharomyces cerevisiae* supplementation on ruminal parameters and milk production of ruminants. *Journal of Dairy Science*, 92(4), 1620-1632.

**Dias, A. L. G., Freitas, J. A., Micai, B., Azevedo, R. A., Greco, L. F. & Santos, J. E. P.** (2018). Effect of supplemental yeast culture and dietary starch content on rumen fermentation and digestion in dairy cows. *Journal of Dairy Science*, 101(1), 201-221.

**Doležal, P., Doležal, J. & Trínáctý, J.** (2005). The effect of *Saccharomyces cerevisiae* on ruminal fermentation in dairy cows. *Czech Journal of Animal Science*, 50(11), 503-510.

**Enjalbert, F., Garrett, J. E., Moncoulon, R., Bayourthe, C. & Chicoteau, P.** (1999). Effects of yeast culture (*Saccharomyces cerevisiae*) on ruminal digestion in non-lactating dairy cows. *Animal Feed Science and Technology*, 76, 195-206.

**Erasmus, L. J., Botha, P. M., & Kistner, A.** (1992). Effect of yeast culture supplement on production, rumen fermentation, and duodenal nitrogen flow in dairy cows. *Journal of Dairy Science*, 75(11), 3056-3065.

**Harrison, G. A., Hemken, R. W., Dawson, K. A., Harmon, R. J., & Barker, K. B.** (1988). Influence of addition of yeast culture supplement to diets of lactating

cows on ruminal fermentation and microbial populations. *Journal of Dairy Science*, 71(11), 2967-2975.

**Hristov, A. N.** (2013). Diet formulation as an effective tool for mitigating nitrogen excretion in dairy systems. *Advances in Animal Biosciences*, 4(s1), 15-18.

**Hristov, A. N., Varga, G., Cassidy, T., Long, M., Heyler, K., Karnati, S. K. R., Corl, B., Hovde, C. J., & Yoon, I.** (2010). Effect of *Saccharomyces cerevisiae* fermentation product on ruminal fermentation and nutrient utilization in dairy cows. *Journal of Dairy Science*, 93(2), 682-692.

**Kamra, D. N., Chaudhary, L. C., Agarwal, N., Singh, R., & Pathak, N. N.** (2002). Growth performance, nutrient utilization, rumen fermentation and enzyme activities in calves fed on *Saccharomyces cerevisiae* supplemented diet. *Indian Journal of Animal Sciences*, 72(6), 472-475.

**Karr, K. J., McLeod, K. R., Dawson, K. A., Tucker, R. E., & Mitchell, G. E.** (1991). Influence of yeast culture and/or monensin on nitrogen flow and rumen fermentation in sheep. In: *Biotechnology in the Feed Industry* (Lyons T. P., ed.). Alltech Tech. Publ., Nicholasville, KY.

**Kebreab, E., France, J., Mills, J. A. N., Allison, R., & Dijkstra, J.** (2002). A dynamic model of N metabolism in the lactating dairy cow and an assessment of impact of N excretion on the environment. *Journal of Animal Science*, 80(1), 248-259.

**Lascano, G. J., Heinrichs, A. J., & Tricarico, J. M.** (2012). Substitution of starch by soluble fiber and *Saccharomyces cerevisiae* dose response on nutrient digestion and blood metabolites for precision-fed dairy heifers. *Journal of Dairy Science*, 95(6), 3298-3309.

**Leicester, H. C. vdW., Robinson, P. H., & Erasmus, L. J.** (2016). Effects of two yeast based direct fed microbials on performance of high producing dairy cows. *Animal Feed Science and Technology*, 215, 58-72.

**Li, Y., Shen, Y., Niu, J., Guo, Y., Pauline, M., Zhao, X., Li, Q., Cao, Y., Bi, C., Zhang, X., Wang, Z., Gao, Y., & Li, J.** (2021). Effect of active dry yeast on lactation performance, methane production, and ruminal fermentation patterns in early-lactating Holstein cows. *Journal of Dairy Science*, 104(1), 381-390.

**McAllister, T. A., Beauchemin, K. A., Alazeh, A. Y., Baah, J., Teather, R. M., & Stanford, K.** (2011). Review: The use of direct fed microbials to mitigate pathogens and enhance production in cattle. *Canadian Journal of Animal Science*, 91, 193-211.

**Miller-Webster, T., Hoover, W. H., Holt, M., & Nocek, J. E.** (2002). Influence of yeast culture on ruminal microbial metabolism in continuous culture. *Journal of Dairy Science*, 85(8), 2009-2014.

**Moallem, U., Lehrner, H., Livshitz, L., Zachut, M., & Yakoby, S.** (2009). The effects of live yeast supplementation to dairy cows during the hot season on production,

feed efficiency, and digestibility. *Journal of Dairy Science*, 92(1), 343-351.

**Newbold, C. J., McIntosh, F. M., & Wallace, R. J.** (1998). Changes in the microbial population of a rumen-simulating fermenter in response to yeast culture. *Canadian Journal of Animal Science*, 78, 241-244.

**Newbold, C. J., Wallace, R. J., Chen, X. B., & McIntosh, F. M.** (1995). Different strains of *Saccharomyces cerevisiae* differ in their effects on ruminal bacterial numbers in vitro and in sheep. *Journal of Animal Science*, 73, 1811-1819.

**Palladino, R. A., Olmeda, M. F., Juliano, N., Bargo, F., & Ipharraguerre, I. R.** (2022). Daily fecal pH pattern and variation in lactating dairy cows. *JDS Communications*, 3, 106-109.

**Panda, A. K., Singh, R., & Pathak, N. N.** (1995). Effect of dietary inclusion of *Saccharomyces cerevisiae* on growth performance of crossbred calves. *Journal of Applied Animal Research*, 7(2), 195-200.

**Perdomo, M. C., Marsola, R. S., Favoreto, M. G., Adesogan, A., Staples, C. R., & Santos, J. E. P.** (2020). Effects of feeding live yeast at 2 dosages on performance and feeding behavior of dairy cows under heat stress. *Journal of Dairy Science*, 103(1), 325-339.

**Phesatcha, K., Phesatcha, B., Wanapat, M., & Cherdthong, A.** (2022). The effect of yeast and roughage concentrate ratio on ruminal pH and protozoal population in Thai native beef cattle. *Animals*, 12, 53.

**Pinloche, E., McEwan, N., Marden, J., Bayourthe, C., Auclair, E., & Newbold, J.** (2013). The effects of a probiotic yeast on the bacterial diversity and population structure in the rumen of cattle. *PLoS ONE*, 8(7), e67824.

**Piva, G., Belladonna, S., Fusconi, G., & Sicbaldi, F.** (1993). Effects of yeast on dairy cow performance, ruminal fermentation, blood components, and milk manufacturing properties. *Journal of Dairy Science*, 76(9), 2717-2722.

**Rotz, C. A.** (2004). Management to reduce nitrogen losses in animal production. *Journal of Animal Science*, 82(E. Suppl.), E119-E137.

**Tristant, D., & Moran, C. A.** (2015). The efficacy of feeding a live probiotic yeast Yea-Sacc<sup>®</sup>, on the performance of lactating dairy cows. *Journal of Applied Animal Nutrition*, 3(e12), 1-6.

**Weiss, W. P.** (2004). Factors affecting manure excretion by dairy cows. In: *Proceedings of Cornell Nutrition Conference, Oct. 19-20*, Syracuse, NY, 11-20.

**Wiedmeier, R. D., Arambel, M. J., & Walters, J. L.** (1987). Effect of yeast culture and *Aspergillus oryzae* fermentation extract on ruminal characteristics and nutrient digestibility. *Journal of Dairy Science*, 70(10), 2063-2068.

**Williams, P. E. V., & Newbold, C. J.** (1990). Rumen probiosis: The effects of novel microorganisms on rumen fermentation and ruminant productivity. In: *Recent Advances in Animal Nutrition* (Haresign W., Cole D. J. A., eds.), Butterworths, London, England.

**Williams, P. E., Walker, A., & MacRae, J. C.** (1990). Rumen probiosis: The effects of addition of yeast culture (viable yeast *Saccharomyces cerevisiae* plus growth medium) on duodenal protein flow in wether sheep. *Proceedings of Nutrition Society*, 49(2), 128A.

**Wohlt, J. E., Corcione, T. T., & Zajac, P. K.** (1998). Effect of yeast on feed intake and performance of cows fed diets based on corn silage during early lactation. *Journal of Dairy Science*, 81(5), 1345-1352.

**Wohlt, J. E., Finkelstein, A. D., & Chung, C. H.** (1991). Yeast culture to improve intake, nutrient digestibility, and performance by dairy cattle during early lactation. *Journal of Dairy Science*, 74(4), 1395-1400.

**Xiccato, G., Schiavon, S., Gallo, L., Bailoni, L., & Bittante, G.** (2005). Nitrogen excretion in dairy cow, beef and veal cattle, pig, and rabbit farms in Northern Italy. *Italian Journal of Animal Science*, 4(Suppl.3), 103-111.

**Yoon, I. K., & Stern, M. D.** (1996). Effects of *Saccharomyces cerevisiae* and *Aspergillus oryzae* cultures on ruminal fermentation in dairy cows. *Journal of Dairy Science*, 79(3), 411-417.